CO₂ assimilation in the chemocline of Lake Cadagno is dominated by a few types of phototrophic purple sulfur bacteria

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Abstract

Lake Cadagno is characterized by a compact chemocline that harbors high concentrations of various phototrophic sulfur bacteria. Four strains representing the numerically most abundant populations in the chemocline were tested in dialysis bags in situ for their ability to fix CO₂. The purple sulfur bacterium Candidatus ‘Thiodictyon syntrophicum’ strain Cad16T had the highest CO₂ assimilation rate in the light of the four strains tested and had a high CO₂ assimilation rate even in the dark. The CO₂ assimilation of the population represented by strain Cad16T was estimated to be up to 25% of the total primary production in the chemocline. Pure cultures of strain Cad16T exposed to cycles of 12 h of light and 12 h of darkness exhibited the highest CO₂ assimilation during the first 4 h of light. The draft genome sequence of Cad16T showed the presence of cbbL and cbbM genes, which encode form I and form II of RuBisCO, respectively. Transcription analyses confirmed that, whereas cbbM remained poorly expressed throughout light and dark exposure, cbbL expression varied during the light–dark cycle and was affected by the available carbon sources. Interestingly, the peaks in cbbL expression did not correlate with the peaks in CO₂ assimilation.

Introduction

Lake Cadagno (Switzerland) is a crenogenic meromictic lake characterized by a narrow chemocline with high concentrations of sulfates; steep gradients of oxygen, sulfide, and light; and a turbidity maximum that correlates with large concentrations of bacteria of up to 10⁷ cells mL⁻¹ (Tonolla et al., 1999, 2004, 2005). Studies in other similarly stratified lakes have confirmed the importance of phototrophic sulfur bacteria to the carbon and sulfur cycles (Pimenov et al., 2003; García-Cantizano et al., 2005; Dimitriu et al., 2008; Casamayor et al., 2012). In Lake Cadagno, the chemocline is typically found at a depth of about 12 m and is characterized by the presence of a dense community of anaerobic phototrophic sulfur bacteria, including purple sulfur bacteria (PSB) of the genera Chromatium, Lamprocystis, Thiocystis and Thiodictyon and green sulfur bacteria (GSB) of the genus Chlorobium (Tonolla et al., 1999, 2004, 2005). From 1998 to 2004 the total concentration of phototrophic sulfur bacteria increased from 4.9 (± 0.2) to 14 (± 1.4) 10⁶ cells mL⁻¹, primarily due to a remarkable rise in the population of the GSB Chlorobium clathrati-forme (Decristophoris et al., 2009; Gregersen et al., 2009). Previous reports have shown that despite its small volume (approximately 10% of the lake), the chemocline of Lake Cadagno is responsible for up to 40% of the total inorganic carbon photo-assimilation (Camacho et al., 2001). Interestingly, significant rates of CO₂ assimilation were also found to occur during the night, indicating that primary production also relies on mechanisms other than photosynthesis. Chemoautotrophic bacteria such as those belonging to the Thiobacillus genus are often responsible for CO₂ fixation in the absence of light (Martinez et al., 1983) but they have not been detected in Lake Cadagno (Tonolla et al., 1999, 2003; Bosshard et al.,...
Form III is found only in some Archaea (each of 50 in the Lake Cadagno chemocline (Musat et al., 1986; de Wit & van Gemerden, 1987). Previous nano-scale secondary-ion mass spectrometry (nanoSIMS) has shown that the inorganic carbon uptake is highly variable among three populations of phototrophic sulfur bacteria in the Lake Cadagno chemocline (Musat et al., 2008).

Green sulfur bacteria assimilate CO₂ via the reverse tricarboxylic acid (rTCA) cycle, whereas PSB assimilate CO₂ via the Calvin-Benson-Bassham cycle in which ribulose-1,5-bisphosphate carboxylase/oxygenase (RuBisCO; EC 4.1.1.39) is a key enzyme. RuBisCO is found in most autotrophic organisms, as well as in eukaryotic algae and plants (Ellis, 1979; Tabita, 2005; Mulkidjianian et al., 2006). There are four major forms of RuBisCO (I–IV) known in nature, distinguishable by their primary amino acid sequences (Badger & Bek, 2008; Tabita et al., 2008). Forms I, II and III of RuBisCO contain catalytically active amino acid residues that are necessary for carboxylation and oxygenation. In contrast, form IV of RuBisCO (also known as the RuBisCO-like protein), which is found in several non-phototrophic bacteria (e.g. Bacillus subtilis) and GSB, is thought to be incapable of CO₂ fixation because it lacks several conserved residues considered essential for RuBisCO carboxylase activity (Saito et al., 2009). Form I, the most abundant form of RuBisCO, is found in many eukaryotes and bacteria and is a multimeric enzyme comprising eight CbbL subunits of 50–55 kDa that are associated with eight CbbS subunits of 12–16 kDa (Schneider et al., 1992). Form II is found in certain Proteobacteria and consists of 2–8 CbbM subunits (each of 50–55 kDa) depending on the microorganism. Form III is found only in some Archaea and comprises large subunit homomers in arrangements of either 2 (L₂) or 5 (L₅) subunits (Tabita et al., 2008).

The aim of this study was to determine the CO₂ assimilation of the various populations of phototrophic sulfur bacteria that are found in the chemocline of Lake Cadagno. To do this, we compared the CO₂ assimilation rates in pure cultures of strains isolated from Lake Cadagno and representing the four numerically most abundant phototrophic sulfur bacteria populations. The pure cultures were cultivated, and subsequently pulsed with NaH¹⁴CO₃, in dialysis bags positioned at a depth of 12 m (described as in situ conditions). In addition, the expression of the RubisCO cbbL (Form I) and cbbM (Form II) genes in pure cultures of the strain Candidatus ‘Thiodictyon syntrophicum’ Cad16T were determined over time during light–dark cycles under laboratory conditions (described as in vitro conditions). Strain Cad16T was chosen as the model organism because it has been shown to represent one of the most prominent CO₂-assimilating populations in the lake both in the light and in the dark.

**Materials and methods**

**Media and growth conditions**

The major characteristics of the strains used in this study are described in Table 1. PSB were grown in Pfennig’s medium I (Eichler & Pfennig, 1988), and GSB were grown in Pfennig’s medium II (Biebl & Pfennig, 1979), both of which contain 0.25 g of KH₂PO₄ L⁻¹, 0.34 g of NH₄Cl L⁻¹, 0.5 g of MgSO₄·7H₂O L⁻¹, 0.25 g of CaCl₂·2H₂O L⁻¹, 0.34 g of KCl L⁻¹, 1.5 g of NaHCO₃ L⁻¹, 0.02 mg of vitamin B₁₂ L⁻¹ and 0.5 mL of trace element solution SL12 L⁻¹ for PSB and SL10 for GSB (see Supporting Information, Table S1). The media were prepared in 2-L bottles using a flushing gas composition of 80% N₂ and 20% CO₂ according to Widdel &

<table>
<thead>
<tr>
<th>Name</th>
<th>Characteristic physiology</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Candidatus ‘Thiodictyon syntrophicum’ strain Cad16T</td>
<td>Purple sulfur bacterium; cells are ovoid-shaped, 1.4–2.4 μm in size, Gram-negative; anaerobic or microaerophilic growth; CO₂ fixation by the Calvin-Benson-Bassham (CBB) cycle</td>
<td>Peduzzi et al. (2012)</td>
</tr>
<tr>
<td>Thioctis chemoclnalis strain CadH11T</td>
<td>Purple sulfur bacterium; cells are ovoid-shaped, 2.3–3.6 μm in size, Gram-negative; anaerobic or microaerophilic growth; CO₂ fixation by the Calvin-Benson-Bassham (CBB) cycle</td>
<td>Peduzzi et al. (2011)</td>
</tr>
<tr>
<td>Lamprocystis purpurea strain CadA31</td>
<td>Purple sulfur bacterium; cells are ovoid-shaped, 1.9–2.3 μm in size, Gram-negative; anaerobic or microaerophilic growth; CO₂ fixation by the Calvin-Benson-Bassham (CBB) cycle</td>
<td>Eichler &amp; Pfennig, (1988)</td>
</tr>
<tr>
<td>Chlorobium clathratiforme strain Cad4DE</td>
<td>Green sulfur bacterium; cells are rod-shaped, 0.3–1.2 μm in size, Gram-negative; strictly anaerobic growth; CO₂ fixation by the reverse tricarboxylic acid (rTCA) cycle</td>
<td>Tonolla et al. (2004)</td>
</tr>
</tbody>
</table>
Bak (1992) and were reduced by the addition of 1.10 mM Na₃S·9H₂O and adjusted to a pH of approximately 7.0. All cultures were incubated at room temperature (20–23 °C) and subjected to a light/dark photoperiod of 12 h with a light intensity of approximately 6 μE m⁻² s⁻¹ that was generated with incandescent 60 W bulbs. All media promoted autotrophic growth except for the mixotrophic medium used for the cbbL/cbbM mRNA expression analysis, to which 2 mM of sodium acetate was added. Concentrations of sulfide in the cultures were measured daily and adjusted to about 1 mM throughout the experiments. The growth was followed by measuring the optical density of culture aliquots at a wavelength of 650 nm (OD₆₅₀) using a UV/VIS Spectrometer Lambda 2S (Perkin-Elmer Inc, Waltham, MA). All biochemical analyses were performed on cells taken from exponentially growing cultures with an OD₆₅₀ of c. 0.6, which corresponds to c. 1.0 × 10⁷ cell mL⁻¹.

**Strain identification, total cell count and viability tests**

Bacterial populations in the chemocline were identified using fluorescent in situ hybridization (FISH) with species-specific Cy3-labeled oligonucleotides (Table 2) in 1-μL aliquots of paraformaldehyde-fixed water samples (n = 3) spotted onto gelatin-coated slides [0.1% gelatin, 0.01% KCr(SO₄)₂] (Glöckner et al., 1996). The hybridizations were performed as described by Zarda et al. (1997). The slides were treated with Citifluor AF1 (Citifluor Ltd., London, UK) and examined by epifluorescence microscopy using filter sets F31 (AHF Analysentechnik, Tübingen, Germany; D360/40, 400DCLP, and D460/50 for DAPI) and F41 (AHF Analysentechnik; HQ535/50, Q565LP, and HQ610/75 for Cy3). Twenty fields of 0.01 mm² were counted, and cell densities were expressed as the mean number of cells mL⁻¹ (±SE). After the incubation period of 5 weeks in the dialysis bags at a depth of 12 m, the number of viable cells was determined using the Live/Dead BacLight Bacterial Viability kit (Invitrogen – Life Technologies Europe, Zug, Switzerland).

### In situ analysis

Meromictic Lake Cadagno is located in the Piora Valley at 1921 m above sea level in the southern Swiss Alps (46°33′N, 8°43′E). Physicochemical parameters of the water column were determined using a YSI 6000 profiler (Yellow Springs, Inc., Yellow Springs, OH) and included temperature (°C), conductivity (μS cm⁻¹), pH, dissolved oxygen (mg L⁻¹), redox potential (mV) and turbidity (FTU, formazine turbidity unit) (Table S1). Two LI-193SA spherical quantum sensors (LI-COR Ltd, Lincoln, NE) were used to determine the percentage of transmittance down to the chemocline of photosynthetically active radiation (%PAR-light). Sulfide concentrations were measured using Cline’s reagent of water samples fixed with ZnCl₂ (Cline, 1969).

In situ ¹⁴CO₂ assimilation analyses were performed as follows: each of the four selected strains was grown separately under laboratory conditions until the cultures reached an OD₆₅₀ of 0.6. This initial period of growth lasted for approximately 10 days due to the relatively long generation times, ranging from 110 to 130 h of cultures. Then, 750 mL of these pure bacterial cultures were sealed in 60-cm-long dialysis bags (inflated diameter of 62.8 mm; Karl Roth GmbH Co. KG, Karlsruhe, Germany) and acclimatized for 5 weeks (from 8 August to 12 September 2007) in the chemocline at a depth of 12 m. With pores 25–30 Å in diameter, the dialysis bags allowed for the free diffusion of molecules smaller than 20 kDa while preventing contamination of incubated cultures by environmental bacteria (Siméon et al., 2001, 2006; Corno & Jürgens, 2006; Blom et al., 2010; Lindström & Ostman, 2011). After this 5-week pre-incubation period, the cultures were transferred from the dialysis bags into independent 100-mL sealed bottles with 125 μL of NaH¹⁴CO₃ for primary production measurements. To limit the exposure of the bacterial culture to atmospheric oxygen, the

**Table 2. Cy3-labeled oligonucleotide probes used in this study for FISH counting**

<table>
<thead>
<tr>
<th>Probe</th>
<th>Target</th>
<th>Sequence (5′→3′)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>S453F</td>
<td>Candidatus ‘Thiodictyon syntrophicum’ strain Cad16²</td>
<td>CCCTCATGGGATTACCCACAAAGGGC (40%)</td>
<td>Tonolla et al. (1999)</td>
</tr>
<tr>
<td>S453H</td>
<td>Thioctys chs ocmlinalis strain CadH11²¹</td>
<td>GACCGAACGTTAACGGCCCGCCTT (10%)</td>
<td>Tonolla et al. (2005)</td>
</tr>
<tr>
<td>Apur453</td>
<td>Lamprocystis purpurea strain CadA31</td>
<td>TGCCACGTTATATCCCAACGAC (40%)</td>
<td>Tonolla et al. (1999)</td>
</tr>
<tr>
<td>Chlc190</td>
<td>Chlorobium clathratiforme strain Cad4DE</td>
<td>GGCAGAACACCATGCCGATTG (20%)</td>
<td>Tonolla et al. (2005)</td>
</tr>
</tbody>
</table>
cultures were transferred in a CO₂-saturated environment. Due to the short exposure time, neither the pH nor the dissolved inorganic carbon (DIC) in the sample were modified by the sublimation of CO₂ (Data S2).

**Measurements of radioactive inorganic carbon assimilation**

The inorganic carbon fixation was evaluated in parallel in one opaque (dark) and one transparent (light) sealed 100-mL bottle. The inorganic carbon assimilation activity was measured using radioactive ¹⁴C isotope (NEC-086S NaH¹⁴CO₃; 1 mCi; 8.40 mCi/mmol; 1-mL ampoules with specific activities of 20 μCi mL⁻¹; Perkin Elmer, Schwarzenbach, Switzerland). A total of 125 μL NaH¹⁴CO₃ solution, with an activity of 0.05 μCi mL⁻¹, was added to each 100-mL bottle. All bottles were completely filled with either a bacteria-free sample from the chemoclone (filtrated with 0.22-μm filter), a chemoclone water sample (12 m) or with bacterial samples from the dialysis bags. Each sample was analysed in triplicate. The filled bottles were placed at a depth of 12 m in the chemoclone and incubated for 4 h (from 12:00 to 16:00 h). Subsequently, the total radioactive carbon that was assimilated was measured by reverse transcription-quantitative PCR (qRT-PCR) analysis using a Light Cycler® instrument (Roche Applied Science) and the QuantiTect® SYBR® Green RT-PCR kit (Qiagen) according to the manufacturer’s instructions. Two contigs were deposited in the NCBI public database with the accession numbers JQ780325 and JQ780326.

**Draft genome sequencing**

The draft genome of *Candidatus ‘Thiodictyon syntrophicum’* strain Cad16ᵀ was determined by pyrosequencing in the laboratory of Dr. S. C. Schuster (Z. Liu, K. Vogl, N.-U. Frigaard, L. P. Tomsho, S. C. Schuster and D. A. Bryant, pers. commun.) at the Genomics Core Facility of Pennsylvania State University. Paired-end reads from GX-20 FLX Titanium chemistries were assembled into 1352 primary contigs. Two contigs predicted to code for *cbbL* and *cbbM* were deposited in the NCBI public database under the respective accession numbers JQ780325 and JQ780326.

**RNA extraction**

Cells of strain Cad16ᵀ were grown in the laboratory under previously described conditions. At specific times, 0.5 mL of the cultures (c. 0.5 × 10⁷ cells) were mixed with 1.0 mL of RNeasy Protect Bacteria Reagent from the RNeasy Protect Mini Kit (Qiagen) and processed according to the manufacturer’s instructions. Total cellular RNA was purified with the RNeasy Mini Kit (Qiagen), resuspended in RNase-free H₂O and stored at −80 °C to minimize degradation.

**Reverse transcription quantitative PCR**

The levels of *cbbL* and *cbbM* gene transcription were measured by reverse transcription-quantitative PCR (qRT-PCR) analysis using a Light Cycler® instrument (Roche Applied Science) and the QuantiTect® SYBR® Green RT-PCR kit (Qiagen) according to the manufacturer’s instructions. Two contigs were deposited in the NCBI public database with the accession numbers JQ780325 and JQ780326. As templates to design the specific primer pairs to determine the expression of *cbbL* and *cbbM* genes, the primers used were cbbL-F (5′-cttcggtcctgggcc-3′), cbbL-R (5′-gcacctgctacatct-3′) and cbbM-F (5′-caggccagatttctctgc-3′), cbbM-R (5′-ttacctgcactacatgc-3′) for *cbbL* and *cbbM*, respectively. The qRT-PCR reaction mixture contained the following components for both primer sets: 10 μL of SYBR Green buffer, 0.4 μL of both primers (0.2 μM primer final concentration), 5 μL of RNA template, 0.2 μL of qRT-PCR mix, 0.1 μL of UNG and DNase free water (Qiagen) to a total volume of 20 μL. The qRT-PCR mix was loaded onto the LightCycler® system, and the reaction conditions were set up as follows: (1) 20-min reverse transcription at 50 °C; (2) 15-min initial activation step at 95 °C; (3) 60 reaction cycles: 0 s at 81 °C, 15 s at 94 °C, 20 s at 63 °C and 10 s at 72 °C; (4) melting curve: 0 s at 95 °C, 15 c at 60 °C and 0 s at 99 °C; (5) 2 min at a 40 °C cooling step. To monitor the progress of the PCR reaction, a fluorescence signal was measured at a wavelength of 530 nm at the end of each elongation phase. A relative quantification analysis was performed using the LightCycler® 4.1, which enabled a comparison of the ratio of the target mRNA in each sample to the reference standard curve. The standard curve was calculated using serial dilutions of *cbbL* or *cbbM* mRNA amplified from cultured bacteria with the previous sets of primers.

**Results**

**Physicochemical and biological properties of Lake Cadagno**

Sample collection and *in situ* experiments were carried out on 12 September 2007. The main physicochemical parameters of the water column on this day, including ATP values, are shown in Fig. 1. The water column was stratified into three zones: the oxic mixolimnion (from the surface to approximately 11 m), the anoxic chemoclone...
(at approximately 12 m), and the anoxic, sulfide-containing monimolimnion (from approximately 13 m down to the sediment). The transition zone from the oxic to the anoxic water layer was also revealed by the redox potential (ORP) by a shift from positive to negative values at approximately 11 m. The turbidity maximum and the ATP values corresponded with a dense microbial community of \(1 \times 10^7\) cells mL\(^{-1}\) that populated the chemocline and consisted mostly of GSB and PSB (as determined by FISH). The GSB \(C.\ clathratiforme\) was the most abundant species with \(8.3 \times 10^6\) cells mL\(^{-1}\) (72.9% of the total bacterial concentration) at 12 m. The PSB \(Candidatus\ Thiodictyon\ syntrophicum\) strain Cad16\(^T\), \(Lamprocystis\ purpurea\) and \(Thiocystis\ chemocinalis\) strain CadH11\(^T\) were clearly less abundant, with \(1.3 \times 10^5\) cells mL\(^{-1}\) (1.2% of total), \(4.9 \times 10^4\) cells mL\(^{-1}\) (0.4% of total) and \(1.9 \times 10^4\) cells mL\(^{-1}\) (0.2% of total), respectively.

**CO\(_2\)** assimilation in the chemocline of Lake Cadagno

To measure the inorganic carbon assimilated in the chemocline and the impact of light on this phenomenon, a water sample was collected from a depth of 12 m. After 4 h of incubation (from 12:00 to 16:00 h on a sunny day), the concentration of fixed \(CO_2\) was determined to be 297 (± 52) ng of \(^{14}C\) mL\(^{-1}\) in the presence of light and 231 (± 98) ng of \(^{14}C\) mL\(^{-1}\) in the dark (see Fig. 2a).

In parallel, the uptake of inorganic carbon was also measured using pure cultures of the four main autotrophic strains from Lake Cadagno (Table 1), which were pre-incubated for equilibration for 5 weeks at a depth of 12 m in dialysis bags as described above. After the equilibration period, the dialysis bags were retrieved, and more than 99% of the cultures consisted of live cells.
As shown in Fig. 2b, Candidatus ‘T. syntrophicum’ Cad16T and L. purpurea CadA31 were clearly the most efficient CO₂-fixing strains, with levels of 0.61 (± 0.11) and 0.39 (± 0.09) pg of ¹⁴C cell⁻¹ in the light, respectively, whereas C. clathratiforme Cad4DE and T. chemoclinis CadH11T showed low to negligible activities of 0.001 (± 0.0002) and 0.04 (± 0.02) pg of ¹⁴C cell⁻¹, respectively. Interestingly, both strains Cad16T and CadA31 showed a strong carbon uptake of 0.41 (± 0.09) and 0.24 (± 0.07) pg of ¹⁴C per cell, respectively, in opaque bottles, indicating that both strains retained the ability to fix CO₂ for at least 4 h in the dark.

As reported in Fig. 2a, a total of 528 ng (297 light + 231 dark) of CO₂ (arbitrarily set as 100%) was fixed by microorganisms found in each millimeter of the chemocline after 4 h of incubation in the presence or absence of light. Given the respective rates per cell of carbon assimilation measured for pure cultures of strains Cad4DE, CadH11T, Cad16T and CadA31 when incubated in the dialysis bags, we attempted to estimate the respective contributions of each population represented by the strains (in % of the total) to the global pool of carbon fixed in the lake. This estimation included the concentration of each of the populations in the chemocline as determined by FISH (see Table 3). In this model, the population represented by Candidatus ‘T. syntrophicum’ Cad16T appeared to be the most important contributor among the four populations tested, because it assimilated as much as 25.9% of all the hypothetical carbon fixed within the chemocline (light and dark). However, the primary CO₂-assimilating organism(s) within the chemocline, responsible for 66.1% of the carbon fixation, still remain(s) unknown.

**CO₂ assimilation of Candidatus ‘T. syntrophicum’ Cad16T grown under controlled laboratory conditions**

To gain an insight into the dynamics of CO₂ assimilation in the organism that was determined to be most efficient at C-fixation, a time course experiment was carried out under laboratory conditions. Cells of Cad16T were grown in Pfennig’s Medium to c. 0.6 OD₆₅₀ under light/dark photoperiods of 12 h. Thereafter, NaH¹⁴CO₃ was added every 4 h, to 50-mL aliquots that were further incubated for 4 h prior to quantification of ¹⁴CO₂. As shown in Fig. 3, CO₂ assimilation reached a maximum (0.89 ± 0.03 pg of ¹⁴C per cell) during the first light period (from 07:00 to 11:00 h) and decreased rapidly once cells were exposed to the dark (0.14 ± 0.03 pg of ¹⁴C per cell between 19:00 and 23:00 h), ending at a minimum assimilation activity of 0.04 ± 0.003 pg of ¹⁴C cell⁻¹ between 23:00 and 03:00 h. The period between 11:00 and 15:00 h, with an in vitro assimilation of 0.53 ± 0.08 pg of ¹⁴C per cell, is comparable to the in situ activity of 0.61 ± 0.11 pg of ¹⁴C cell⁻¹ measured during the time frame from 12:00 to 16:00 h in the light. Similarly, the ¹⁴CO₂ assimilation of 0.41 ± 0.09 pg of ¹⁴C
per cell between 19:00 and 23:00 h corresponded to the in situ 14CO2 assimilation in the dark. The level of sulfide was monitored during the experiment, but no correlation with other parameters was found. The maximum sulfide level was at 15:00 h after 8 h of light, and the minimum was at 3:00 h after 8 h of dark.

Expression of cbbL and cbbM genes in Candidatus ‘T. syntrophicum’ Cad16T in the presence or absence of an organic carbon source

Two putative genes coding for the RuBisCO form I (cbbL) and form II (cbbM) were found in the genome annotation of strain Cad16T (Data S3). To establish which form of RuBisCO is the most important for CO2 fixation in strain Cad16T, the transcript levels of cbbL and cbbM were determined in anoxic autotrophically or anoxic mixotrophically grown cells. When cultures reached an OD650 of 0.6, aliquots were collected at 2–4 h intervals, and total RNA was prepared as described in the Materials and methods section. Figure 4 shows the levels of cbbL and cbbM transcripts measured using qRT-PCR from total cellular RNA samples. In cells growing autotrophically, cbbL was actively transcribed, with two distinct peaks of expression at 15:00 h (12.4/0.01 pg of mRNA per 10^6 cells) and 03:00 h (15.4/0.01 pg of mRNA per 10^6 cells). In contrast, under mixotrophic conditions (2 mM of acetate), the expression of cbbL remained approximately 10-fold lower than the expression under autotrophic conditions. Although the expression was lower, under mixotrophic conditions, a high level of expression was recorded at 15:00 and 03:00 h, with 0.88/0.004 and 0.69/0.004 pg of mRNA per 10^6 cells, respectively. Moreover, the sampling point showing the minimal expression of cbbL was at 23:00 h, with 0.12/0.014 pg of mRNA per 10^6 cells corresponding to the minimal expression under autotrophic conditions, with 3.21/0.036 pg of mRNA per 10^6 cells.

Expression of the locus that codes for form II RuBisCO remained constant and at a low level throughout the experiment for both incubation conditions. The expression of cbbM was approximately 5-fold higher in autotrophic compared to mixotrophic conditions. However, the maximum level of expression, at 09:00 h (0.06 ± 0.009 pg of mRNA per 10^6 cells) in autotrophic and at 19:00 h (0.017 ± 0.002 pg of mRNA per 10^6 cells) in mixotrophic growing conditions, and the minimum level of expression, at 07:00 h (0.022 ± 0.006 pg of mRNA per 10^6 cells) in autotrophic and at 11:00 h (0.004 ± 0.001 pg of mRNA per 10^6 cells) in mixotrophic growing conditions, were independent of each other.

Table 3. Total estimated contributions to CO2 fixation by selected groups of organisms in Lake Cadagno (phototrophic populations are represented by specific strains)

<table>
<thead>
<tr>
<th>Strains</th>
<th>Populations in the chemocline* (10^4 cells mL^-1)</th>
<th>Levels of CO2 fixation</th>
<th>Total CO2 fixed (ng C mL^-1)</th>
<th>% of total</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Light + Dark† (pg C per 10^6 cells)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>C. clathratiforme Cad4DE</td>
<td>831</td>
<td>8 + 4</td>
<td>11</td>
<td>2.0</td>
</tr>
<tr>
<td>T. chemoclinis CadH11T</td>
<td>2</td>
<td>450 + 40</td>
<td>1</td>
<td>0.2</td>
</tr>
<tr>
<td>Candidatus ‘T. syntrophicum’ Cad16T</td>
<td>13</td>
<td>6100 + 4100</td>
<td>137</td>
<td>25.9</td>
</tr>
<tr>
<td>L. purpurea CadA31</td>
<td>5</td>
<td>3900 + 2400</td>
<td>31</td>
<td>5.8</td>
</tr>
<tr>
<td>Others</td>
<td>289</td>
<td>Not determined</td>
<td>Not determined</td>
<td>66.1</td>
</tr>
<tr>
<td>Chemocline sample†</td>
<td>1140</td>
<td></td>
<td>528†</td>
<td>100.0</td>
</tr>
</tbody>
</table>

*Cells for each strain count in the chemocline (12 m) by FISH the same day of the experiment showed in Fig. 2 (12 September 2007).
†Values shown in Fig. 2B.
‡Total cell count in the chemocline (12 m) by DAPI the same day of the experiment showed in Fig. 2 (12 September 2007).
§Experimentally determined by the sum of values shown in Fig. 2A (see text).
their CO2 assimilation in the chemocline at a depth of 20 m (Fig. 2). Among the strains representing the four most abundant populations of phototrophic sulfur bacteria in the chemocline, *Candidatus* ‘*T*. syntrophicum’ Cad16T was the most effective CO2-fixing organism, in both the presence and the absence of light. Surprisingly, the most abundant phototrophic sulfur bacterial population in the chemocline, the GSB *C. clathratiforme* constituting 73% of total bacterial cells (Tonolla et al., 2005; Decristophoris et al., 2009; Gregersen et al., 2009) was the least efficient in terms of CO2 assimilation per cell, with values that were approximately 100-fold lower than those measured for the *Candidatus* ‘*T*. syntrophicum’ Cad16T. This finding confirms previous results from the Lake Cadagno chemocline photoprophic community, which showed that *C. clathratiforme* was clearly a less effective assimilator of CO2 than were *Chromatium okenii* and *L. purpurea* (Musat et al., 2008). In our study, *L. purpurea* CadA31 showed high rates of CO2 assimilation similar to *Candidatus* ‘*T*. syntrophicum’ Cad16T (Fig. 2b); moreover, these two species are also phylogenetically closely related (Garrity et al., 2007). Musat et al. (2008) also showed that *C. okenii* (0.3% of the chemocline’s population) contributed up to 70% of the total inorganic carbon fixed during daylight. A similar estimation is proposed by us in Table 3, taking in account both light and dark CO2 assimilation, in which it can be seen that *Candidatus* ‘*T*. syntrophicum’ Cad16T was the main assimilator measured, being responsible for about 25.9% of the total assimilation. However, the four most abundant populations of phototrophic sulfur bacteria under study were responsible for only 33.9% of the total CO2 fixed in the chemocline, and the remaining 66.1% was not identified. According to Musat et al. (2008), we can speculate that the remaining 66.1% of CO2 assimilation in the chemocline is carried out by *C. okenii*. Unfortunately, *C. okenii* could not be grown as a pure culture in the laboratory and therefore the CO2 assimilation by this population could not be estimated using our approach.

The chemocline of Lake Cadagno showed high rates of carbon assimilation, not only in the presence of light of c. 297 ng of 14C mL−1 but also in the dark at c. 231 ng of 14C mL−1 (Fig. 2a). Similarly high CO2 assimilation rates in the dark were also reported for other stratified lakes, such as the Spanish karstic lakes (Casamayor et al., 2008, 2012; Casamayor, 2010), Lake Kinneret in Israel (Hadas et al., 2001) and Big Soda Lake in Nevada (Cloern et al., 1983). In Lake Cadagno, the populations represented by *Candidatus* ‘*T*. syntrophicum’ Cad16T and *L. purpurea* CadA31 were responsible for approximately one-third (28.7%) of the dark assimilation (data not shown). Prior to the increase in the population of *C. clathratiforme* (Tonolla et al., 2005; Decristophoris et al., 2009; Gregersen et al., 2009), analysis of the CO2 assimilation in the chemocline showed high rates of CO2 assimilation in the dark. At this time *C. okenii* was among the dominant populations of phototrophic sulfur bacteria (Schanz et al., 1998; Camacho et al., 2001). This might indicate that PSB, and in particular *C. okenii*, are the types of organisms mostly responsible for these high dark fixation rates. This speculation remains to be confirmed by further analysis. There is also the possibility that a different metabolic pathway (such as chemolithoautotrophy) in unknown organisms may contribute to the dark carbon fixation activity in this ecosystem (Yngve Borshieim et al., 1985; Jorgensen et al., 1991; Shively et al., 1998), to the fermentation of stored glycogen (Gfeller & Gibbs, 1984; Habicht et al., 2011), and to redox potential-balancing metabolisms (McKinlay & Harwood, 2010) or other unknown metabolisms (Martinez-Garcia et al., 2011). To put it briefly, the chemocline of Lake Cadagno showed a high activity of dark CO2 assimilation that is not completely understood.

**Discussion**

The chemocline of the meromictic Lake Cadagno harbors a complex microbial ecosystem in which phototrophic sulfur bacteria play a key role in inorganic carbon fixation (Camacho et al., 2001). Using dialysis bags that allowed bacterial cultures to remain pure while exposing and equilibrating them to their natural environment (Simek et al., 2001, 2006; Corno & Jürgens, 2006; Blom et al., 2010; Lindström & Ostman, 2011), we were able to assess their CO2 assimilation in the chemocline at a depth of 12 m (Fig. 2). Among the strains representing the four most abundant populations of phototrophic sulfur bacteria in the chemocline, *Candidatus* ‘*T*. syntrophicum’ Cad16T was the most effective CO2-fixing organism, in both the presence and the absence of light. Surprisingly, the most abundant phototrophic sulfur bacterial population in the chemocline, the GSB *C. clathratiforme* constituting 73% of total bacterial cells (Tonolla et al., 2005; Decristophoris et al., 2009; Gregersen et al., 2009) was the least efficient in terms of CO2 assimilation per cell, with values that were approximately 100-fold lower than those measured for the *Candidatus* ‘*T*. syntrophicum’ Cad16T. This finding confirms previous results from the Lake Cadagno chemocline photoprophic community, which showed that *C. clathratiforme* was clearly a less effective assimilator of CO2 than were *Chromatium okenii* and *L. purpurea* (Musat et al., 2008). In our study, *L. purpurea* CadA31 showed high rates of CO2 assimilation similar to *Candidatus* ‘*T*. syntrophicum’ Cad16T (Fig. 2b); moreover, these two species are also phylogenetically closely related (Garrity et al., 2007). Musat et al. (2008) also showed that *C. okenii* (0.3% of the chemocline’s population) contributed up to 70% of the total inorganic carbon fixed during daylight. A similar estimation is proposed by us in Table 3, taking in account both light and dark CO2 assimilation, in which it can be seen that *Candidatus* ‘*T*. syntrophicum’ Cad16T was the main assimilator measured, being responsible for about 25.9% of the total assimilation. However, the four most abundant populations of phototrophic sulfur bacteria under study were responsible for only 33.9% of the total CO2 fixed in the chemocline, and the remaining 66.1% was not identified. According to Musat et al. (2008), we can speculate that the remaining 66.1% of CO2 assimilation in the chemocline is carried out by *C. okenii*. Unfortunately, *C. okenii* could not be grown as a pure culture in the laboratory and therefore the CO2 assimilation by this population could not be estimated using our approach.

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Our long-term goal is to understand the microbial basis of CO₂ fixation in Lake Cadagno in the light and in the dark. For this purpose, we have established laboratory protocols to reproducibly cultivate and examine strains that we estimate represent the vast majority of the bacteria found in Lake Cadagno’s chemocline – with the notable exception of C. okenii. The ability to maintain these bacteria as pure cultures allowed us to study the molecular mechanisms responsible for primary production. Candidatus ‘T. syntrophicum’ Cad16 T was used as a model organism to better understand the dynamics of carbon fixation in the chemocline of the Lake Cadagno. While rates of CO₂ assimilation in the light were comparable between in vitro (0.53 ± 0.08 pg of ¹⁴C per cell) and in situ (0.61 ± 0.11 pg of ¹⁴C per cell) conditions, this was not the case for rates in the dark. Indeed, the CO₂ assimilated in vitro in the first 4 h of dark, at c. 0.14 pg ¹⁴C per cell, was approximately threefold lower than the value observed in situ in the dialysis bags c. 0.41 pg ¹⁴C per cell. The absence of oxygen and organic substrates (e.g. acetate) in the autotrophic Pfennig medium I suggested two possible explanations. The absence of oxygen in vitro may prevent the possibility of chemolithotrophic metabolism, in a way similar to that reported for the PSB Thiocapsa roseopersicina (de Wit & van Gemerden, 1987; Schaub & van Gemerden, 1994; Ende et al., 1996). However, the high rate of dark fixation normally combines different metabolic processes more than simple chemolithotrophy (Zopfi et al., 2001). In a highly reduced environment such as the chemocline of the Lake Cadagno, the ability of cells to maintain the redox balance can be very important. Recently, it was proposed that CO₂ fixation is important not only as a primary production mechanism but also as an electron-accepting process in mixotrophic organisms (Richardson et al., 1988; Hallenbeck et al., 1990; Tavano et al., 2005). The Calvin cycle in the purple non-sulfur bacterium Rhodobacter palustris was shown to re-oxidize nearly half of the reduced cofactors generated during the conversion of acetate to biomass, revealing that CO₂ fixation plays a major role in cofactor recycling (McKinlay & Harwood, 2010). The difficulty in cultivating organisms in vitro highlights the problems encountered in attempting to dissect a complex microbial ecosystem such as the chemocline of Lake Cadagno and underscores the need for techniques that would enable studies in situ, such as the dialysis bags.

High rates of CO₂ assimilation in the early hours of light exposure (Fig. 3) have also been observed in the diurnal cycles of cyanobacteria, and this often coincided with strong expression of RuBisCO-encoding genes (Pichard et al., 1996; Wyman, 1999; Paul et al., 2000). In the draft genome sequence of Candidatus ‘T. syntrophicum’ Cad16 T, we identified two RuBisCOs genes, a form I cbbL and a form II cbbM (Data S3). Transcription analyses confirmed that cbbL expression was confirmed at 15:00 and 03:00 h (Fig. 4). This suggests that a cyclic mechanism, possibly synchronized by light, regulates the expression of cbbL in autotrophic conditions. In contrast, the presence of a suitable carbon source (e.g. acetate) in the growth medium caused reduced transcription of cbbL. Similar observations were reported for other PSBs such as Allochromatium vinosum (Valle et al., 1988; Kobayashi et al., 1991). The absence of regulation of the transcription of cbbM under the tested growth conditions may suggest that this process is not regulated by environmental factors and therefore is constitutive. Differential expression of forms I and II of RuBisCO was also reported in other phototrophic bacteria, for which it was proposed that in presence of a reduced carbon source (e.g. acetate), CbbM functions primarily as a terminal electron acceptor involved in maintaining the redox balance of the cell (Wang et al., 1993; Yoshizawa et al., 2004; Badger & Bek, 2008; Joshi et al., 2009; Laguna et al., 2010).

In conclusion, our results show that of the four strains isolated from the chemocline of Lake Cadagno and for which we established in situ and in vitro growth conditions, PSB Candidatus ‘Thiodictyon syntrophicum’ strain Cad16 T exhibited the highest CO₂ assimilation activity, both in presence of light and in the dark. Laboratory experiments using pure cultures of Candidatus ‘T. syntrophicum’ Cad16 T grown in an autotrophic medium allowed us to detect that the maximal CO₂ assimilation rate occurred between 07:00 and 11:00 h, during the first 4 h of light (Fig. 2). However, this maximal activity did not correlate with the expression of any RuBisCO genes (cbbL and cbbM) (Fig. 3). In fact, RuBisCO form I cbbL showed two peaks of expression at 15:00 and 03:00 h, which occur more than 4 h before and 4 h the main assimilation period. The measured expression of RuBisCOs genes suggested that form I cbbL is dependent on environmental conditions, such as light exposure and carbon sources, whereas the form II cbbM gene appears to be constitutively expressed.

**Acknowledgements**

Financial support for this project was provided by the University of Geneva, the Institute of Microbiology and State of Ticino, and the Swiss National Science Foundation (grant no. 31003A-116591). The authors are indebted to P. Decristoforitis, A. P. Caminada, N. Ruggeri and C. Strambio-De-Castillia for technical and moral support. Furthermore, we are grateful to S. Bianchi, R. Bernasconi and M. Molinari, Institute for Research in Biomedicine (IRB), in Bellinzona (Switzerland), for their technical
References


CO₂ fixation by the PSB Thiodictyon syntrophicum Cad16⁷


**Supporting Information**

Additional Supporting Information may be found in the online version of this article:

- **Data S1.** Recipe of trace elements SL10 and SL12.
- **Data S2.** Dissolved inorganic carbon (DIC) and pH from dialysis bags.
- **Data S3.** Genome analysis.
- **Table S1.** Physical parameters of the Lake Cadagno of the 12 September 2007.
- **Fig. S1.** Specific FISH counting of GSB (white) and PSB (black) compared to the total prokaryotic cells counted by DAPI (grey) at different depths of Lake Cadagno during the day of the 14CO₂ assimilation analysis from the cultures pre-incubated in dialysis bags (*in situ*, September 12, 2007).